

Kikuyu (*Pennisetum clandestinum*) intake determined by alkanes administered in a xantham gum suspension

J. Mann[#] and P.G. Stewart

KwaZulu-Natal Department of Agriculture and Environmental Affairs,
Private Bag X 9059, Pietermaritzburg 3200, South Africa

Abstract

The alkane method of measuring herbage intake in ruminants was compared with actual intake determined by transponder-controlled feed gates (Calan gates). Kikuyu (*Pennisetum clandestinum*) intakes of yearling bulls (Holstein-Friesland and Hereford) were measured for a period of seven days. No significant effect of breed on intake was found. Herbage intake calculated from the alkane technique (mean of morning and afternoon faecal sampling times) compared favourably with actual intake with an average intake of 6.25 kg DM/day (93 g DM/kg W^{0.75}/day) (CV 22%). However, a significant effect in the faecal sampling times on intake was observed. Herbage intake determined from alkane concentrations in the faecal samples that were collected in the morning underestimated actual intake by 11%, while intake calculated from faecal alkane concentrations of samples that were collected in the afternoon overestimated actual intake by 8%. It is hypothesised from this study that the dosing of the alkane (dotriacontane) should be every 12 hours when administered in a xantham gum suspension.

Keywords: Alkanes, intake, kikuyu, ruminant

[#]Corresponding author. E-mail: joanne.mann@dae.kzntl.gov.za

Introduction

The alkane technique (Dove & Mayes, 1991; 1996) has been widely used to estimate individual herbage intake (and digestibility) of ruminants. Intake is determined by the ratio of the orally administered synthetic alkane (dotriacontane) to the naturally occurring alkane (trtriacontane). Although herbage intake measured by the alkane method has an error of less than 5% (Dove & Mayes, 1991) when compared with actual intake, most of these validation studies were done on either perennial ryegrass (*Lolium perenne*) or natural pasture (mostly *Festuca* species). Various studies have been conducted using the alkane technique to measure intake on kikuyu (*Pennisetum clandestinum*), but either actual intake was not measured (Horne, 1996; Reeves *et al.*, 1996b; Fushai, 1997) or time of faecal sampling was not considered (Reeves *et al.*, 1995). Various methods of alkane administration to the ruminant have been used. These include: controlled release devices (Reeves *et al.*, 1995), gelatine capsules (Reeves *et al.*, 1996b; Dove *et al.*, 2000), pellets (Hameleers & Mayes, 1998) and a suspension (Horne, 1996; Marais *et al.*, 1996; Fushai, 1997). The studies that have used the suspension prepared by Marais (Marais *et al.*, 1996) did not measure actual intake of kikuyu. The aim of this study was to assess how the alkane intake estimate (using the suspension by Marais *et al.*, 1996) would compare with actual intake on kikuyu.

Material and Methods

The dry matter (DM) intake of nine yearling bulls (five Hereford and four Holstein-Friesland) was measured in a Calan gate facility (transponder-controlled feed gate; described by Broadbent *et al.*, 1970) to compare with alkane intake estimates over a period of seven days (following 30 days of adaptation).

Animals had *ad libitum* access to cut kikuyu grass and a mineral supplement. The grass was cut daily to a residual height of 40 mm with a sickle bar mower. The herbage was further chopped with a forage harvester to a length of *ca.* 25 mm to prevent wastage. The herbage was weighed in and out of feed troughs daily.

The animals were orally administered 1 g of dotriacontane daily at 6:00. The dotriacontane was coated onto dried milled kikuyu (using a rotary evaporator) which was then suspended in a 0.4% xantham gum solution, (Marais *et al.*, 1996). Faecal samples (to be analysed for dotriacontane (C₃₂) and tritriacontane (C₃₃) concentration) were collected from the rectum of each animal daily at 6:00 and 18:00.

Kikuyu grass samples were analysed for crude protein (AOAC, 1995; method 990.03), acid detergent fibre (AOAC, 1995; method 973.18), neutral detergent fibre (Van Soest *et al.*, 1991), ash (AOAC, 1990; method 942.05), calcium (AOAC, 1995; method 968.08) and phosphorus (AOAC, 1990; method 965.17).

The alkane concentration of the coated grass (administered orally), kikuyu grass and faecal samples was analysed for dotriacontane (C₃₂) and tritriacontane (C₃₃) concentrations, according to Marais' procedure (Marais *et al.*, 1996) with some modifications (J.P. Marais, 1997, KwaZulu-Natal Department of Agriculture and Environmental Affairs, Pietermaritzburg. Pers. comm.) which are summarized in Table 1. Samples were not analysed in duplicate as in the procedure by Marais *et al.* (1996) because of cost. The internal standard was changed from tetratriacontane (C₃₄), which was used in Marais *et al.* (1996) to hexatriacontane (C₃₆).

Table 1 Modifications made to Marais' procedure (Marais *et al.*, 1996) (J.P. Marais, 1997, Pers. comm.)

Measurement	Marais <i>et al.</i> (1996)	Current study
Coating of dotriacontane onto solid support		
Milled grass particle size	0.5 to 1.0 mm	0.5 mm
Rotary evaporator temperature during C ₃₂ coating onto grass support	40 °C	60 °C
Coated grass sieved to remove lumps	1 mm sieve	0.5 mm sieve
Dotriacontane coated grass analysis		
Quantity of petroleum ether (80-100 °C BP) added to coated grass sample	30 mL	40 mL
Quantity of solvent added to silica gel column to elute alkanes	10 mL hexane	25 mL petroleum ether (60-80 °C BP)
Grass analysis		
Quantity of grass analysed	-	1.5 g
Faecal analysis		
Quantity of solvent added to silica gel column to elute alkanes	10 mL hexane	25 mL petroleum ether (60-80 °C BP)
Quantity of hexane in which the extracted alkanes were dissolved (before injected into gas chromatograph)	0.3 mL	0.7 mL

Grass and faecal alkane (dotriacontane and tritriacontane) concentrations were substituted in the equation by Dove & Mayes (1991) to calculate intake:

$$\text{Intake } \left(\frac{\text{kg DM}}{\text{d}} \right) = \frac{\frac{F_i}{F_j} \times D_j}{H_i - \left(\frac{F_i}{F_j} \times H_j \right)}$$

where

- D_j = daily dose of the even-chain alkane (C₃₂)
- F_i = faecal concentrations of the odd-chain alkane (C₃₃)
- H_i = herbage concentrations of the odd-chain alkane (C₃₃)
- F_j = faecal concentrations of the even-chain alkane (C₃₂)
- H_j = herbage concentrations of the even-chain alkane (C₃₂).

Statistical differences between treatment means for intake were determined from analysis of variance tables (Genstat 5 (Release 3.2) statistical program) using least significant differences.

Results and Discussion

The chemical analysis of the herbage fed is given in Table 2. The low crude protein, high neutral detergent fibre and high acid detergent fibre values of the herbage on offer, when compared with the studies by Reeves *et al.* (1996a) and Dugmore (1998) indicate that it was either very mature kikuyu (high stem fraction) or nitrogen fertilization may have been lacking (T.J. Dugmore, 2000, KwaZulu-Natal Department of Agriculture and Environmental Affairs, Pietermaritzburg. Pers. Comm.)

Table 2 Mean chemical composition (g/kg DM) of the daily kikuyu on offer (n = 8) compared with two studies from the literature (Reeves *et al.*, 1996a; Dugmore, 1998)

Measurement	Herbage	Reeves <i>et al.</i> (1996a)	Dugmore (1998)
Crude protein (g/kg DM)	87.6	207.5	200
±s.e.	±4.6		
Neutral detergent fibre (g/kg DM)	709.3	602.5	650
±s.e.	±13.6		
Acid detergent fibre (g/kg DM)	390.7	230.9	350
±s.e.	±8.8		
Ash (g/kg DM)	84.9		
±s.e.	±4.3		
Calcium (g/kg DM)	4.1	3.1	2.4
±s.e.	±0.2		
Phosphorus (g/kg DM)	2.2	3.1	3.3
±s.e.	±0.1		

Actual and alkane estimated kikuyu intake is presented in Table 3. No significant effect of breed on intake was measured; therefore, the data was pooled.

Table 3 Actual and calculated intake (alkane technique) of cut kikuyu herbage of yearling bulls

Measurement	Actual intake (n = 9)	Herbage intake calculated by the C ₃₃ -C ₃₂ alkane pair			CV %
		Mean (n = 9)	Morning ^s (n = 9)	Afternoon ^s (n = 9)	
Intake (kg DM/d)	6.28 ^b	6.21 ^b	5.61 ^a	6.81 ^c	23
±s.e.	±0.24	±0.15	±0.17	±0.22	
Intake (g DM/kg W ^{0.75} /d)	93.6 ^b	92.9 ^b	83.7 ^a	102.2 ^c	22
±s.e.	±3.16	±2.07	±2.17	± 3.12	

^{a,b,c} Values bearing different superscripts in the same row are significantly different (P < 0.05)

^s Intake determined from faecal samples collected at either 06:00 or 18:00

Although the intake of kikuyu herbage calculated by the alkane technique did not differ from actual intake when the sampling times were combined, significant trends were found when analysed for time of day. Herbage intake calculated from the C₃₃-C₃₂ concentrations in the morning faecal samples resulted in actual intake being underestimated by 11% (P < 0.05) while herbage intake calculated from alkane concentrations in the afternoon faecal samples overestimated actual intake by 8% (P < 0.05). Literature on the alkane method of intake estimation recommended that faecal sampling could be done once a day since the variability of the faecal C₃₃:C₃₂ ratio is less than 5% when expressed as a proportion of the mean (Mayes *et al.*, 1986b). Only nine of the 15 points of the current study fell within the 5% range when expressed as a proportion of the mean. An analysis of variance was conducted on the C₃₃:C₃₂ ratio and confirmed the significant difference (differed by 18%; P < 0.05) between the morning and afternoon C₃₃:C₃₂ ratio and consequently intake estimates.

It is unlikely that the effect of the faecal sampling time (06:00 and 18:00) on intake could be attributed to diurnal variation as the alkane method has been used previously to predict intake accurately (Mayes *et al.*, 1986a, b; Dillon & Stakelum, 1989; Reeves *et al.*, 1995).

Reeves *et al.* (1996b) compared the alkane method with the rising plate meter and the standard energy requirements in reverse to determine kikuyu intake of dairy cows. Although Reeves *et al.* (1996b) concluded that the alkane method is a “direct and precise method of measuring the intake of individual cows grazing tightly-managed kikuyu pasture”, actual intake was not measured. Furthermore, Reeves *et al.* (1996b) did not find a significant difference in intake determined by the morning (06:00) and afternoon (14:00) sampling times (eight hour interval). Hard-shell gelatine capsules containing alkanes were embedded in powdered cellulose, were administered twice a day in Reeves’ study.

In the study by Hameleers & Mayes (1998), dairy cows were administered alkane pellets (containing dotriacontane) twice a day and faecal samples were collected twice a day after milking (times were not stipulated). The herbage consumed was perennial ryegrass (*Lolium perenne*) and white clover (*Trifolium repens*). No significant effect ($P > 0.05$) was detected in intake determined by morning and afternoon sampling times and actual intake.

A study (Dove *et al.*, 2000) where dosing (alkane capsules) and faecal collection times were carried out twice a day (09:00 and 16:00; seven hour interval) using sheep, similarly found no difference in alkane concentration (hence intake) between twice a day faecal grab samples and total faecal collection (determined by faecal bags). The herbage on offer was temperate pasture.

Although the actual kikuyu intake measured by Reeves *et al.* (1995) did not differ significantly from intake calculated by the C₃₂-C₃₃ alkane pair, time of faecal sampling was not investigated. Faecal samples were collected twice daily but were pooled across days. The success of the alkane technique in the study by Reeves *et al.* (1995) might be attributed to the method of alkane administration, which was a controlled release device.

Besides the study by Reeves *et al.* (1995) no other studies on kikuyu, which also compared the alkane technique with actual intake were found in the literature. The findings of the current paper might possibly be explained by the dosing schedule as opposed to the faecal sampling schedule. From the alkane studies mentioned in the literature, the dosed alkane (usually dotriacontane) was administered twice a day either in capsule (Reeves *et al.*, 1996b; Dove *et al.*, 2000), in pellet form (Hameleers & Mayes, 1998) or continuously by a controlled release device (Reeves *et al.*, 1995). The variability of the C₃₃:C₃₂ ratio between the morning and afternoon faecal sampling times of the current study could be due to the dotriacontane concentration varying in the rumen. Another dose of dotriacontane is possibly required within a 24 hour period to maintain a continuous flow of dotriacontane through the digestive tract. Although once a day dosing and faecal sampling has been accepted as adequate when using the alkane technique (Mayes *et al.*, 1986b), it can be hypothesised from this study that the dosing of the alkane (dotriacontane) should be every 12 hours when administered in a xantham gum suspension. The faecal sampling schedule requires further investigation.

Conclusion

From this study it is hypothesised that the dosing of the alkane (dotriacontane) should be every 12 hours when administered in a xantham gum suspension, as described by Marais *et al.* (1996).

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